Screening for Charcoal Rot Resistance Under field Environment



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Soybean (Glycine max)

Charcoal rot is a soil borne fungus caused by Macrophomina phaseolina

It causes a seedling blight, root rot or stem rot of at least 500 species of plants including many wild and cultivated species



• *Macrophomina phaseolina* infects a wide range of plant species.

Soybean
Corn
Sorghum
Sunflower
Beans



Environmental Conditions

- Soil Temperatures between 82 95 °F favors the disease
- 2. Favors limited soil Moisture
- 3. It is a nutrient scavenger. It thrives and competes well in nutrient depleted soils.

Agronomic Impact of Charcoal Rot

- Charcoal rot can reduce plant height, root volume, and root weight by more that 50%.
- Damage to the root system is most evident during the pod formation and filling stages.
- Because diseased plants have smaller root systems, seeds tend to be fewer and lighter.
- Diseased plants will mature several weeks earlier, which further contributes to yield loss.

Management Approaches Attempted

1) Crop rotation with a less susceptible crop

- 2) Minimizing plant stress by avoiding excessive seeding rates
- 3) Fertilizing when necessary
- 4) Irrigating to keep soil moisture high during pod development periods
- 5) Efficacy of fungicides applications to seed and soil to reduce or inhibit fungal germination and infection.

Genetic Resistance

There is no commercial cultivar that is completely resistant to charcoal rot but a tolerant cultivar has been identified and released as a germplasm.

However, still new sources of resistance are needed.

Number of Soybean Lines Tested Each year for Charcoal Rot Evaluation for Resistance Between 2002-2009



Screening For Resistance

Evaluation of breeding lines
Commercial RR cultivars
Conventional (non-RR) cultivars
Germplasms

For Screening for Charcoal Rot Resistance 1. Inoculum Increase



2. Field Innoculation





Symptoms Early Foliar Symptoms



Symptoms



Responses of different soybean lines to charcoal rot





Ultimate Damage of Charcoal Rot



Comparison of Disease Measurements

- Colony forming Unit (CFU)
- Stem Severity Rating
- Percent height of discoloration
- Foliar Symptom taken @ R7 growth stage
- Foliar Symptom taken @ R1, R3, R5, R7 and calculate the Area Under the disease progress curve (AUDPC)

In developing a method for assessment

We used five methods24 cultivars in MG 3, 4 and 5.

Symptoms Early Foliar Symptoms

- 1. Foliar @ R7
- 2. Foliar @ R1, R3, R5, R7



Disease Assessment

_3 & 4. **R7 - R8**



5. Root & Stem Sections

ground







Sclerotia of *Macrophomina phaseolina* on lower stem & root sections



Comparison of Five Methods for Charcoal Assessment





Relationship Between SSR and CFU of *M. Phaseolina*





Additional Resistant Lines

DT99-16864, DT99-17483, DT17554
 We are currently working on potential resistant lines:

- Six PI Lines
- Several SCN and conventional selections

We now know that there is genetic variability in soybean germplasm for reaction to charcoal rot.

> We have identified more soybean lines that are moderately resistant to charcoal rot.

Identified soybean lines with higher level of resistance. These lines, require further validation and testing over the next two years.

□Ultimately these lines will be used by public and private breeders for cultivar development.



Challenges in Screening for Resistance

Macrophomina phaseolina

Base on a preliminary study: >We know that there is genetic variability within isolates of M. phaseolina. It is a highly variable and quite heterogeneous, with isolates differing in microsclerotia size. Some isolate produce pycnidia and other not.





Macrophomina phaseolina isolates from different locations and hosts

ISOLATE	LOCATION	HOST
TN5	Ames, TN	Soybean (<i>Glycine max</i> (L.) Merr.)
TN146	Neosho, MO	Soybean (<i>Glycine max</i> (L.) Merr.)
TN261	Milan, TN	Soybean (<i>Glycine max</i> (L.) Merr.)
TN272	Stoneville, MS	Soybean (<i>Glycine max</i> (L.) Merr.)
TN280A†	Jackson, TN	Soybean (<i>Glycine max</i> (L.) Merr.)
TN280B	Jackson, TN	Soybean (<i>Glycine max</i> (L.) Merr.)
TN291A	Jackson, TN	Snapbean (<i>Phaseolus vulgaris</i> L.)
TN291B	Jackson, TN	Snapbean (<i>Phaseolus vulgaris</i> L.)
TN292	Milan, TN	Sunflower (<i>Helianthus annuus</i> L.)
TN293A	Jackson, TN	Pumpkin (<i>Cucurbita pepo</i> L.)
TN293B	Jackson, TN	Pumpkin (<i>Cucurbita pepo</i> L.)
TN294	Milan, TN	Maize (Zea mays L.)
TN295A	Dyer Co., TN	Sorghum (<i>Sorghum bicolor</i> (L.) Moench)
TN295B	Dyer Co., TN	Sorghum (<i>Sorghum bicolor</i> (L.) Moench)
TN296	Jackson, TN	Cotton (<i>Gossypium hirsutum</i> L.)
TN305	Madison, WI	Soybean (<i>Glycine max</i> (L.) Merr.)
TN314	Columbus, Cherokee Co., KS	Soybean (<i>Glycine max</i> (L.) Merr.)
TN377	Dunklin Co., MO	Soybean (<i>Glycine max</i> (L.) Merr.)
TN378	Stoddard Co., MO	Soybean (<i>Glycine max</i> (L.) Merr.)
TN379	New Madrid Co., MO	Soybean (<i>Glycine max</i> (L.) Merr.)
TN4 (I-4280)	Jackson, TN	Soybean (<i>Glycine max</i> (L.) Merr.)
TN410	Jackson, TN	Sunflower (<i>Helianthus annuus</i> L.)
TN411	Milan, TN	Sunflower (<i>Helianthus annuus</i> L.)
TN413	Jackson, TN	Maize (Zea mays L.)

Dendrogram showing the relationships between isolates. (Arias, et al. 2010)



Screening for Resistance

- Based on this finding, our future focus will be to characterize the soybean isolates even further and identify isolates that are more aggressive.
- Use the aggressive pathotype (s) to identify resistance.

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